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In Silico Analysis and Computational Assessment of Potential Inhibitors of SARS-Cov-2 Main Protease

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Abstract

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) outbreak in China has caused so many deaths with a significant number of confirmed cases around the world. This virus's highly contagious nature has raised the scientific community's concern to find a cure to treat coronavirus disease (COVID-19) caused by the viral infection. In this study 1615, FDA approved ligand structures were analyzed to identify the best possible therapeutic inhibitor(s) for SARS-CoV-2 main protease. Upon sequential computational experiments, Lindane (Induced Fit Docking; IFD Score: -594.02 Kcal/mol), Gluconolactone (IFD Score: -585.77 Kcal/mol), and Mitoxantrone (IFD Score: -582.33 Kcal/mol) were found to be the best inhibitors of SARS-CoV-2 main protease. Then these compounds were analyzed in different post-screening experiments where they were also observed to perform well. This study will contribute to the current efforts of scientific society to secure treatment for COVID-19. However, further in vivo and in vitro experiments might be required to properly validate the findings of this study.

Keywords: Coronavirus; COVID-19; Deaths; Inhibitors; Main Protease

INTRODUCTION

Coronavirus is a type of pathogen that causes potentially deadly diseases in mammals and birds [1]. In humans, they may cause severe to mild respiratory, enteral, hepatic, or neurologic diseases with symptoms of fever, cough, and shortness of breath [2]. They are enveloped singlestranded, positive-sense RNA viruses. It is currently known as the largest RNA genome and is phenotypically and genotypically diverse [3].

Coronavirus resides in the family Coronaviridae (generacoronavirus and torovirus) in the order nidovirales [4]. They are widely spread in bats worldwide and found in many other species such as birds, dogs, pigs, mice, cats, and humans [5]. Members of the coronavirus family possess a large genome that ranges from 27-32 kb. This coronavirus genome encodes for 5/ replicase polyprotein containing two reading frames ORFs 1a and 1b, which in order, encodes for all the enzymes necessary for viral RNA replication [6]. According to serology and genome phylogeny, coronaviruses are classified into four genera, termed Alpha, Beta, Delta, and Gramma coronavirus [7]. So far, seven human coronaviruses have been identified, containing two alpha CoVs (HCoV-229E and HCoV-NL63) and five beta CoVs (HCoV-OC43, HCoV-HKU1, severe acute respiratory syndrome CoV (SARS-CoV), middle east respiratory syndrome CoV (MERS-CoV) and most recent SARS-CoV-2 [8].

In the 1960s [9], two human coronaviruses- human coronavirus 229E (HCoV- 229E) and human coronavirus OC43 (HCoV-OC43) were known to infect humans [10] in the lower respiratory tract [11]. After this, a third human coronavirus, SARS-CoV, was discovered responsible for severe acute respiratory syndrome [12]. It was first identified in November 2002, in China's Guangdong region [13,14]. The world health organization (WHO) declared that the disease was spread to 29 areas worldwide in 2003. Eight thousand ninety-eight individuals were infected, and among them, 774 death cases were observed [15]. After the SARS epidemic,

the two other human coronaviruses, HCoV-NL63 and HCoV-HKU1 were discovered quickly. On 23 September 2012, the WHO found a new coronavirus- middle east respiratory syndrome coronavirus (MERS-CoV). In 2012 and December 2019, 2465, MERS-CoV infection cases were confirmed, and 850 deaths were reported from 27 countries to the WHO [10-16].

However, these two highly pathogenic coronaviruses (SARS-CoV and MERS-CoV) caused global epidemics with terrifying morbidity and mortality. In December 2019, another human coronavirus outbreak, causing the COVID-19, was recorded in Wuhan City, Hubei Province, China. According to epidemiological studies, this outbreak was associated with Wuhan's seafood market [17,18]. Based on phylogenetic analysis of the complete viral genome (29,903 nucleotides), it was demonstrated that SARS-CoV-2 was most nearly related (89.1% nucleotides similarity) to the SARS-like coronavirus [19].

And the protein sequences of SARS-CoV-2 main protease (CMP) and SARS-CoV main protease have 96.1% sequence identity (Figure 1).

Viral main protease processes other viral precursor proteins for proper functioning, and therefore blocking their activity is an effective strategy in developing antiviral drugs [22,23]. In this study, a total of 1615 FDA approved compounds were docked against SARS-CoV-2 main protease. The sequential computational analysis led to identifying the best three compounds that were then utilized in another post-screening study (Figure 2).



| (A) | (B) |
|--------|---|
| | 96.1% identity in 306 residues overlap; Score: 1600.0; Gap frequency: 0.8% |
| | COVID-19 1 SGFRKMAFPSGKVEGCMVQVTCGTTTLINGLWLDDVVYCPRHVICTSEDMLNPNYEDLLIR SARS-CoV 1 SGFRKMAFPSGKVEGCMVQVTCGTTTLINGLWLDDTVYCPRHVICTAEDMLNPNYEDLLIR |
| | COVID-19 61 KSNHNFLVQAGNVQLRVIGHSMQNCVLKLKVDTANPKTPKYKFVRIQPGQTFSVLACYNK SARS-CoV 61 KSNHSFLVQAGNVQLRVIGHSMQNCLLRLKVDTSNPKTPKYKFVRIQPGQTFSVLACYNK |
| | COVID-19 SARS-CoV 121 SPSGVYQCAMRPNFTIKGSFLNGSCGSVGFNIDVDCVSFCYMHHMELPTGVHAGTDLEGM 121 SPSGVYQCAMRPNHTIKGSFLNGSCGSVGFNIDVDCVSFCYMHHMELPTGVHAGTDLEGM |
| dikney | COVID-19 SARS-CoV 181 FYGPFVDRQTAQAAGTDTTITVNVLAWLYAAVINGDRWFLNRFTTTLNDFNLVAMKYNYE 181 FYGPFVDRQTAQAAGTDTTITLNVLAWLYAAVINGDRWFLNRFTTTLNDFNLVAMKYNYE |
| | COVID-19 SARS-CoV 241 PLTQDHVDILGPLSAQTGIAVLDMCASLKELLQNGMNGRTILGSALLEDEFTPFDVVRQO 241 PLTQDHVDILGPLSAQTGIAVLDMCAALKELLQNGMNGRTILGSTILEDEFTPFDVVRQO |
| W | COVID-19 301 SGVTFQ SARS-CoV 301 SGVTFQ |

[20,21]. (B) Alignment of SARS-CoV-2 and SARS- CoV main protease protein sequences.

MATERIALS AND METHODS

Molecular Docking

Protein preparation

PDB format of SARS-CoV-2 main protease (SMP) (PDB ID: 67U7) was downloaded from Protein Data Bank (www. rcsb.org) in the three-dimensional crystallographic form [24], which has an inhibitor protein attached to its active site. To prepare and process Protein Preparation Wizard's protein structure in Maestro Schrödinger Suite (v11.4) was used. Assigning the bond orders to the structures, hydrogens were added to the heavy atoms. All of the water molecules were erased from the atoms; missing side chains were adjusted in the protein structure backbone using Prime as well as het states were generated with Epik at pH 7 ± 2 [25]. Optimized Potentials for Liquid Simulations force field (OPLS_2005) in the suite was utilized, setting the RMSD (root-mean-square-deviation) to 30 Å to refine and minimize the protein structure. Any extraordinary water under 3H-bonds to non-water was erased in the minimization step.

Ligand preparation

A total of 1615 FDA approved drug (ligand) structures were downloaded in sdf format from ZINC database (https://zinc.docking.org/) [26]. LigPrep wizard of Maestro Schrödinger suite was used to prepare and process these ligand structures [27]. Minimized 3D structures of the ligands were generated using Epik2.2 within pH 7.0 +/- 2.0 in the suite. Finally, minimization was carried out again using the OPLS_2005 force field, which generated maximum 32 possible stereoisomers depending on available chiral centers on each molecule.

Receptor grid generation

Receptor grid was generated using default Van der Waals radius scaling factor 1.0 and charge cutoff 0.25, which was then subjected to the OPLS_2005 force field for the minimized structure in Glide [28]. The grid generated a cubic box around the active site (co-crystallized reference ligand) of the target molecules. Finally, the grid box dimension was then adjusted to 14 Å ×14 Å×14 Å for docking to be carried out.

Glide standard precision (SP) and extra precision ligand docking

To compare docking parameters, both Extra precision (XP) ligand docking and Standard precision (SP) ligand docking methods are used that are suitable and accurate for small number and the large number of ligand

molecules, respectively [29,30]. To carry out docking of the ligand molecules, Van der Waals radius scaling factor 0.80 and charge cutoff 0.15 were set, which assigned final scores according to the docked ligand's pose within the binding cleft of the receptor molecule. Discovery Studio Visualizer (v4.5) was utilized to analyze the best possible poses and types of ligand-receptor interactions [31].

Prime MM-GBSA rescoring

The ligands were subjected to Molecular mechanicsgeneralized born and surface area (MM- GBSA) rescoring with Prime module of Maestro Schrödinger suite for further evaluation after SP XP ligand docking. MM-GBSA assigns a more accurate scoring function, which in turn improves the overall free binding affinity score upon the reprocessing of the docked ligand to the biological macromolecules by the utilization of an implicit solvent [32-34]. This technique generally combines OPLS molecular mechanics energies (EMM), surface generalized born solvation model for polar solvation (GSGB), and a nonpolar salvation term (GNP) for total free energy (Δ Gbind) calculation. The following equation calculated the total free energy of binding:

 Δ Gbind = Gcomplex – (Gprotein – Gligand), where, G= EMM + GSGB + GNP

Induced fit docking

Ten best compounds with the lowest MM-GBSA scores were selected for further evaluation since it is the more robust scoring method. To obtain more accurate docking result, the chosen ligand molecules were subjected to induced fit docking (IFD) that generates the native poses of the ligands bound to its target [32-35]. After having generated a grid around the receptor's co- crystallized ligand structure, the OPLS_2005 force field was applied again, and the five best ligands were docked rigidly. Van der Waals screening for the receptor (0.70) and ligand (0.50) was set, residues within 2 Å were refined in order to generate the two best possible posses with the standard precision method.

Structure-based Pharmacophore Modeling

Pharmacophore modeling is one of the essential tools used for successful drug discovery. It is usually used for both lead identification and lead optimization and aids in the prompt understanding of 2D or 3D level identity of molecules by schematically depicting the critical elements of molecular recognition [36,37]. 3D structure-based pharmacophore modeling was obtained using LigandScout 4.4.1 Essential [38]. LigandScout autonomously generates pharmacophore features like- hydrogen bond donors, hydrogen bond acceptors, hydrophobic, positive and negative ionizable, aromatic interactions, and 3D geometries of the intended bioactive molecules using an advanced algorithm [39].

ADME/T Prediction

In silico prediction of the ADME/T profile of candidate drug molecules, it helps pharmaceutical industries select the best candidates, which reduces the time and cost of the drug discovery approach [40-42]. ADME/T profile for three selected ligand molecules (Table 3) that performed well in the docking experiment was analyzed using an online based server, i.e., admetSAR 2.0 (http://lmmd. ecust.edu.cn/admetsar2) and pkCSM (http://biosig. unimelb.edu.au/pkcsm/prediction) to predict different pharmacokinetic and pharmacodynamic properties. These are including blood- brain barrier permeability, human abdominal absorption, AMES toxicity, Cytochrome Р inhibitory promiscuity, carcinogenicity, (CYP) mutagenicity, and Caco-2 permeability [43,44].

PASS (Prediction of Activity Spectra for Substances) Prediction

Prediction of Activity Spectra for Substances (PASS) estimates the tentative biological activities of query compounds based on their native chemical structure. PASS predicts the action of a compound based on Structure-Activity Relationship Base (SAR Base), which assumes that the activity of a compound is related to its structure. It works by comparing the 2D structure of a compound relative to another compound having biological activities recorded in the database with almost 95% accuracy [45]. Probable antiviral activities and other intended activities against proteins involved in mediating viral infection of the selected molecules were predicted using PASS online server (http://www.pharmaexpert.ru/passonline/) [46].

DFT (Density Functional Theory) Calculation

DFT calculation was carried out by using minimized ligand structures from LigPrep. This calculation theory uses the Jaguar panel of Maestro Schrödinger Suite, which uses Becke's three- parameter exchange potential as well as Lee-Yang-Parr correlation (B3LYP) theory with 6-31G* basis set [47-50]. Different quantum chemical properties such as surface properties (MO, density, potential) and Multipole moments were calculated along with HOMO (Highest Occupied Molecular Orbital) and LUMO (Lowest Unoccupied Molecular Orbital) energy. Then the global frontier orbital was analyzed as well as hardness (η) and softness (S) of selected molecules were calculated using the following equation as per Parr and Pearson interpretation and Koopmans theorem [51,52]. The DFT calculation result is summarized in Table 6, and the HOMO and LUMO occupation of the ligands is illustrated in.

 $\eta = (HOMOE-LUMOE)/2, S = 1/\eta$

RESULTS

Molecular Docking Study

Among all the ligand molecules used in SP, XP, and MM-GBSA scoring, 10 molecules were selected based on the lowest binding free energies since it is the most rigorous scoring method mentioned earlier (Table 1). Three best performing ligand molecules were selected based on the IFD scores, which is considered a more accurate way of predicting binding poses than SP and XP from 10 selected molecules that showed slight variation between SP and XP docking scores. These three ligand molecules were Lindane, Gluconolactone, and Mitoxantrone (Table 2), and these ligands were found to exhibit the lowest IFD scores (Table 3). As a result, these compounds were utilized for further study, and others were opted out from further consideration.

| Compound ID | SP Docking Score (Kcal/mol) | XP Docking Score (Kcal/mol) | Glide Ligand Efficienc y | Glid e ecoul | Glide evdw | Glide energ Y | MM- GBSA ΔGbind (Kcal/mol) |
|-------------------|--------------------------------|--------------------------------|-----------------------------|-----------------|------------|------------------|--------------------------------|
| ZINC00000381086 0 | -6.749 | -5.328 | -0.078 | -1.34 | - 30.03 3 | - 31.370 | -77.03 |
| ZINC00024520492 4 | -5.874 | -4.751 | -0.313 | - 1.227 | - 21.85 3 | - 23.080 | -64.07 |
| ZINC00002130321 0 | -5.775 | -4.460 | -0.204 | 0.166 | - 33.03 4 | - 32.868 | -62.14 |
| ZINC00000253970 2 | -5.214 | -4.134 | -0.067 | - 4.317 | -41.32 | - 45.637 | -61.05 |
| ZINC00001970230 9 | -5.689 | -5.123 | -0.258 | - 0.352 | -31.93 | - 32.282 | -59.59 |
| ZINC0000089659 5 | -4.893 | -4.477 | -0.213 | -3.45 | - 26.94 3 | - 30.393 | -54.25 |
| ZINC00000379479 4 | -5.683 | -4.172 | -0.264 | - 1.985 | - 23.89 9 | - 25.884 | -53.43 |
| ZINC0000000043 1 | -6.085 | -4.956 | -0.236 | - 5.536 | - 34.79 2 | - 40.328 | -53.18 |
| ZINC00000640973 5 | -5.116 | -4.111 | -0.316 | 0.987 | - 27.32 1 | - 26.334 | -52.77 |
| ZINC00000395288 1 | -5.261 | -3.853 | -0.148 | - 2.888 | - 34.79 6 | - 37.684 | -45.96 |

 Table 1: Result of SP and XP docking and free binding energy calculation.

| Compound ID | Compound Name | MM- GBSA ΔGbind (Kcal/mol) | Glide Gscore (Kcal/mol) | IFD Score (Kcal/mol) | Interacting Amino Acids | Bond Distance (Å) | Type of Interaction | Interaction Category |
|------------------|------------------|----------------------------------|----------------------------|-------------------------|----------------------------|-------------------------|-------------------------|-------------------------|
| | | | | | Cys145 | 4.80 | Alkyl | Hydrophobic |
| | | | | | Met49 | 4.75 | Alkyl | Hydrophobic |
| | | | | | Cys145 | 5.22 | Alkyl | Hydrophobic |
| | | | | | Met49 | 5.30 | Pi-Alkyl | Hydrophobic |
| | | | | | Met165 | 3.67 | Alkyl | Hydrophobic |
| | | | | | His41 | 5.11 | Pi-Pi T Shaped | Hydrophobic |
| | | | | | Met165 | 5.48 | Pi-Alkyl | Hydrophobic |
| | | | | | His41 | 2.92 | Hydrogen Bond | Non- conventional |
| ZINC000245204924 | Lindane | -64.07 | -6.25 | -594.02 | Met165 | 4.92 | Alkyl | Hydrophobic |
| | | | | | His41 | 4.15 | Pi-Alkyl | Hydrophobic |
| | | | | | Arg188 | 3.09 | Chlorine Interaction | Halogen |
| | | | | | His41 | 3.65 | Pi-Alkyl | Hydrophobic |
| | | | | | Met49 | 3.30 | Alkyl | Hydrophobic |
| | | | | | His41 | 4.77 | Pi-Pi T Shaped | Hydrophobic |
| | | | | | Glu166 | 2.27 | Hydrogen Bond | Conventional |
| ZINC000002539702 | Gluconolactone | -61.05 | -3.97 | -585.77 | His41 | 2.78 | Hydrogen Bond | Non- conventional |
| | | | | | Arg188 | 2.59 | Hydrogen Bond | Non- conventional |
| | | | | | Met49 | 4.95 | Pi-Alkyl | Hydrophobic |
| | | | | | Met165 | 2.95 | Pi-Alkyl | Hydrophobic |
| | | | | | His41 | 2.70 | Hydrogen Bond | Conventional |
| | | | | | Ser144 | 2.71 | Hydrogen Bond | Conventional |
| ZINC000003794794 | Mitoxantrone | -53.43 | -3.38 | -582.33 | Asn142 | 2.85 | Hydrogen Bond | Non- conventional |
| | | | | | Gly143 | 2.04 | Hydrogen Bond | Conventional |

Binding mode of lindane with SARS-CoV-2 main protease (SMP)

Lindane is one of the selected best performance showing ligand molecules that interacted with 5 amino acids within the binding pocket and formed a total of 6 interactions when docked with SMP with a better IFD score (-594.02 Kcal/mol) as well as Glide Gscore (-6.25 Kcal/mol) (Table 2). It formed one non-conventional hydrogen bond with His41 amino acid residue at 2.92 Å distance apart, one Pi-Pi T shaped interaction with His41, and one halogen interaction with Arg188 amino acid residue. It also formed additional hydrophobic interactions, i.e., Alkyl and Pi-Alkyl interactions with Cys145, Met49, Met165 amino acid residues within the binding cleft of CMP (Figure 3).

Binding mode of gluconolactone with SARS-CoV-2 main protease (SMP)

Gluconolactone is another best performance showing ligand molecule that generated an IFD score of -585.77 Kcal/mol and Glide Gscore of -3.97 Kcal/mol to dock with SMP and formed total 6 interaction when interacted with 4 amino acids within the binding pocket (Table 2). It formed one conventional hydrogen bond with Glu166 amino acid residue at 2.27 Å distance apart and two non-conventional interactions with His41 and Arg188 amino acid residues at 2.78 and 2.59 Å distance apart,

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| Zinc ID | Compoun d Name | IUPAC Name | Chemical Formula | 2D structure |
|----------------------|--------------------|---|---------------------|--------------|
| ZINC0002452 04924 | Lindane | 1,2,3,4,5,6- hexachlorocyclohexane | C6 H6 Cl6 | |
| ZINC0000025 39702 | Gluconolac tone | (3 <i>R</i> ,4S,5 <i>S</i> ,6 <i>R</i>)-3,4,5-trihydroxy- 6-(hydroxymethyl)oxan-2-one | C6 H10 O6 | |
| ZINC0000037 94794 | Mitoxantro ne | 1,4-dihydroxy-5,8-bis[2-(2- hydroxyethylamino)ethylamino] anthracene- 9,10-dione | C22 H28N4O6 | |



respectively. Additional hydrophobic Pi-Alkyl interaction was also formed with Met49 amino acid residue within the binding cleft of CMP (Figure 4).

Binding mode of mitoxantrone with SARS-CoV-2 main protease (SMP)

Mitoxantrone generated an IFD score of -582.33 Kcal/ mol and Glide Gscore of -3.38 Kcal/mol to dock with SMP and formed a total 6 interactions interacting with 4 amino acids (Table 2). It formed three conventional hydrogen bonds with His41, Ser144, and Gly143 amino acid residue at 2.70, 2.71, and 2.04 Å distance apart. Moreover, Mitoxantrone also created one non-conventional hydrogen bond interaction with Asn142 amino acid residue at 2.85 Å distance apart, respectively. Again, it formed additional hydrophobic Pi-Alkyl interactions with Met165 amino acid residue within the binding cleft of SMP (Figure 4).

Pharmacophore Modeling

The result of pharmacophore modeling is represented in (Figure 4). Lindane was reported to show only hydrophobic features, and Met165 and Met49 amino acid residues were shown to contribute to the bond formation with the pharmacophoric quality. Gluconolactone formed three hydrogen bond donor features and one hydrogen bond acceptor feature, and His41, Met165, and Glu166 amino acid residues were shown to contribute to the bond formation with the pharmacophoric features. Again, Mitoxantrone formed three hydrogen bond donor features within the binding site of CMP.

ADME/T Prediction

ADME/T (absorption, distribution, metabolism, excretion, and toxicity) profile was analyzed for all selected ligand molecules, and the results are summarized in (Table 4). The best three selected ligand molecules

(Lindane, Gluconolactone, and Mitoxantrone) showed high oral bioavailability. Lindane has higher Caco2 cell line permeable capability compare to Gluconolactone

| Table 4: Results of ADME/T tests of best selected ligands. OCT2: |
|--|
| Organic Cation Transporter 2; hERG: Human ether-a-go-go related |
| gene, CYP: Cytochrome P450 |

| Properties | Lindane | Gluconolactone | Mitoxantrone | | | | | | |
|---------------------------------------|---------|----------------|--------------|--|--|--|--|--|--|
| Absorption | | | | | | | | | |
| Human intestinal absorption | High | Low | High | | | | | | |
| Human oral bioavailability | High | High | High | | | | | | |
| Caco-2 permeability | High | Low | Low | | | | | | |
| | Dist | ribution | | | | | | | |
| P-glycoprotein substrate | No | No | Yes | | | | | | |
| P-glycoprotein inhibitor | No | No | No | | | | | | |
| Blood-brain barrier penetration | Yes | Yes | No | | | | | | |
| Metabolism | | | | | | | | | |
| CYP3A4 substrate | No | No | No | | | | | | |
| CYP2C9 substrate | No | No | No | | | | | | |
| CYP2D6 substrate | No | No | No | | | | | | |
| CYP3A4 inhibition | No | No | No | | | | | | |
| CYP2C9 inhibition | No | No | No | | | | | | |
| CYP2D6 inhibition | No | No | No | | | | | | |
| | Exc | retion | | | | | | | |
| Total clearance | 1.053 | 0.678 | 1.424 | | | | | | |
| OCT2 substrate | No | No | No | | | | | | |
| Toxicity | | | | | | | | | |
| AMES toxicity | No | No | Yes | | | | | | |
| Hepatotoxicity | No | No | Yes | | | | | | |
| hERG inhibition | No | No | No | | | | | | |
| Eye irritation | Yes | No | No | | | | | | |
| Acute oral toxicity | 111 | IV | III | | | | | | |



Figure 4: Three dimensional (left) and Two dimensional (right) representation of structure-based pharmacophore modeling between selected ligand molecules and SMP. Hydrogen bond donor, hydrogen bond acceptor, and hydrophobic features are represented as green, red, and yellow.

and Mitoxantrone, respectively. Gluconolactone showed less human intestinal absorption, whereas Lindane Mitoxantrone's absorption rate was high. Plasma membrane protein, P-glycoprotein, was not inhibited by any of the best-selected ligand molecules; Mitoxantrone acted as a substrate. Lindane and Gluconolactone showed the blood-brain permeable capability, but none of the ligands showed the sign as either inhibitor or substrate of CYP3A4, CYP2D6, CYP2C9, and OCT2. None of them was reported to be substrates of OCT2 (Organic Cation Transporter 2). Mitoxantrone was predicted to have Ames toxicity and hepatotoxicity, whereas Lindane was shown to induce eye irritation. On the contrary, Mitoxantrone and Lindane showed type III acute oral toxicity, and Gluconolactone showed type IV.

Pharmacological Activity Prediction

In order to determine the association of ligand molecules with other antiviral activities based on their native structures, their pharmacological activities were predicted, and the result is represented in Table 5. Gluconolactone showed better antiviral activity against different viruses and other actions against different enzymes involved in viral infection mediation. However, Mitoxantrone was predicted to have moderate activity followed by Lindane.

Analysis of Frontier's Orbitals

Detailed HOMO energy, LUMO energy, energy gap (HOMO-LUMO gap), hardness, and softness of the selected best compounds are summarized in Table 6, and the HOMO and LUMO occupation of the ligand molecules is illustrated in (Figures 5 and 6) for each compound. The highest energy gap was predicted for Lindane, and the lowest gap was observed for Mitoxantrone.

According to the energy gap, the stability order of the compounds is: Mitoxantrone>Gluconolactone>Lindane. Along with the HOMO and LUMO energy, each compound's dipole moment was also calculated, and based on the dipole moments, the molecules' stability order is Mitoxantrone>Lindane>Gluconolactone.

DISCUSSION

Due to the utilization of specific algorithms and scoring function as well as the particular pose of ligand-receptor interaction, molecular docking is the most commonly used drug discovery approach by the researchers [53,54]. In this experiment 1615, FDA-approved ligand structures were downloaded from the ZINC database, and then they were subjected to SP, XP, and binding free energy calculation. Out of them, 10 best performing ligand molecules were selected based on their lowest binding energy, reflecting the higher affinity of ligand molecules

Table 5: Result of Pharmacological Activity prediction of selected ligand molecules. Pa>0.7: Compound is very likely to have activity; Pa>0.5:

 Compound is expected to have activity

| A - 41- 241 | Lindane | | Gluconolacton e | | Mitoxantrone | |
|--|---------|-------|-----------------|-------|--------------|-------|
| Activities | Pa | Pi | Ра | Pi | Ра | Pi |
| Antiviral (Picornavirus) | 0.546 | 0.033 | 0.704 | 0.005 | - | - |
| Antiviral (Poxvirus) | 0.373 | 0.034 | 0.678 | 0.011 | 0.348 | 0.040 |
| Simian immunodeficiency virus proteinase inhibitor | 0.525 | 0.016 | 0.466 | 0.029 | 0.286 | 0.138 |
| Antiviral (Adenovirus) | 0.391 | 0.033 | 0.407 | 0.026 | 0.441 | 0.016 |
| Antiviral (Influenza) | 0.265 | 0.116 | 0.687 | 0.006 | 0.334 | 0.071 |
| Antiviral (Herpes) | 0.429 | 0.024 | 0.512 | 0.008 | 0.318 | 0.079 |
| Antiviral (Hepatitis B) | - | - | 0.478 | 0.005 | 0.166 | 0.143 |
| Antiviral | 0.375 | 0.018 | 0.361 | 0.021 | - | - |
| Antiviral (Rhinovirus) | - | - | 0.437 | 0.057 | - | - |
| RNA-directed RNA polymerase inhibitor | 0.438 | 0.030 | 0.484 | 0.013 | - | - |
| RNA directed DNA polymerase inhibitor | 0.503 | 0.009 | 0.408 | 0.017 | 0.213 | 0.076 |
| HIV-2 reverse transcriptase inhibitor | 0.305 | 0.009 | 0.185 | 0.041 | 0.207 | 0.031 |
| 3C-like protease (Human coronavirus) inhibitor | 0.297 | 0.027 | 0.231 | 0.108 | - | - |

Table 6: Result of DFT calculation. The unit of HOMO, LUMO, gap, hardness, and softness are in Hartree, and the unit of dipole moment is in Debye.

| Compound Name | номо | LUMO | Gap | Hardness (η) | Softness (S) | Dipole Moment |
|----------------|----------|----------|---------|--------------|--------------|---------------|
| Lindane | -0.30473 | -0.03623 | 0.2685 | 0.13425 | 7.4487 | 2.2069 |
| Gluconolactone | -0.27343 | -0.02362 | 0.24981 | 0.124905 | 8.0060 | 4.6608 |
| Mitoxantrone | -0.17629 | -0.08850 | 0.08779 | 0.043895 | 22.8003 | 2.0253 |



for their receptors. The selected ligand molecules were again subjected to induced fit docking (IFD) to obtain a more accurate docking result (Table 1). Based on the induced-fit docking experiment scores, Lindane, Gluconolactone, and Mitoxantrone were selected as the best inhibitors of SMP, and their drug-like potentials were then evaluated (Tables 2 and 3). In this study, Lindane, Gluconolactone, and Mitoxantrone showed the lowest binding energies and formed multiple numbers of hydrogen bonds and hydrophobic interactions within the binding site of SMP (Figure 3). These interactions are significant for serving biological purposes, making the ligand-receptor complex more efficient [55,56].

In SARS Coronavirus main protease His41 and Cys145 amino acid residues form the catalytic dyad of the enzyme's active site [57]. In our study, the best-selected ligand molecules formed hydrogen bonds and hydrophobic interaction with either His41 or Cys145 amino acid residue of the active site of CMP. They thus predicted to interfere with the normal function of the protease. Pharmacophore modeling is a fascinating technique for de novo drug designing and lead optimization. Structurebased pharmacophore modeling uses a 3D structure of a macromolecular target or a target-ligand complex

J Virol Infect Dis. (2021) Volume 2 Issue 1 to predict pharmacophoric features of the complex. As a part of the protocol subsequently, structure-based pharmacophore modeling utilizes an assessment of the complementary chemical features of the binding site and their spatial orientation relationships, which results in a pharmacophore model assembly with selected features, i.e., hydrogen bond donors, hydrogen bond acceptors, and hydrophobes [58,59]. The best-docked compounds were reported to have significant hydrogen bond donors, acceptors, and hydrophobic features (Figure 4).

Considering the most concerning issues such as bloodbrain barrier permeability for drugs that usually target cells of the central nervous system (CNS), determining the most efficient route of drugs, the highly absorbed organs or tissues, the ADME/T (absorption, distribution, metabolism, excretion, and toxicity) profile of drugs is analyzed during the development of a potential drug [60,61]. Plasma membrane proteins such as P-glycoproteins play a significant role in the side human body's drug transport mechanism. Human intestinal tissue permeability of a drug is ensured by the Caco2 cell line permeability of that drug [62-64]. Inside the human body, drug interaction, metabolism, and drug excretion outside the body are regulated by the Cytochrome P450 family of enzymes. Inhibition of these enzymes leads to the normal process's impairment and may cause acute drug toxicity, slow clearance, and malfunction of drugs [65,67]. AMES toxicity parameter is used to examine the toxicity endpoint of chemicals under investigation [68,69]. hERG (Human ether-a-go-go related gene) channels are the voltage-gated potassium ion channels that play crucial roles for potassium ion transport through the cell membrane, and this hERG potassium channel can be blocked by structurally and functionally different as well as unrelated drugs that may raise offtarget drug interaction. To minimize the undesirable drug interaction, compounds for hERG channels are screened during the early lead optimization process [70]. The substrates of Renal OCT2 (organic cation transporter 2) are readily excreted through urine, and this transporter is essential for drugs and xenobiotic excretion through the kidney [71]. Lindane was predicted to have better ADME/T profiles than the other two molecules (Table 4).

Pharmacological activity (PASS prediction) refers to the Probability of activity (Pa) and the Probability of inactivity (Pi) of a compound, and the result of the forecast ranges from 0.000 to 1.000. The activity of a compound can only be possible When Pa>Pi [72]. A compound is considered highly active when its Pa value is greater than 0.7 (Pa>0.7), and there is a possibility of that compound being analog to a known pharmaceutical agent is also high. A compound also shows activity when its Pa value is greater than 0.5 but less than 0.7 (0.5<Pa<0.7), but the possibility of being analogue to a known pharmaceutical agent is low, and when a compound has Pa value less than 0.5 (Pa<0.5), it is considered as a less active compound [73]. The selected compounds were analyzed to determine the antiviral activities and activities against proteins and enzymes involved in viral infection, and Gluconolactone was predicted to have better pharmacological activities than other selected molecules (Table 5). The HOMO-LUMO gap defines a compound's stability, and HOMO refers to a constraint portion in a molecule capable of donating electrons, whereas LUMO is responsible for accepting electrons. To determine the stability of the best selected ligand molecules, their HOMO and LUMO energy were analyzed (Figure 5). It is required for a compound to have the highest gap to undergo a chemical reaction more efficiently [74,75]. Lindane was predicted to have a more positive energy gap than other molecules (Table 6). Finally, upon continual computational exploration, Lindane, Gluconolactone, and Mitoxantrone were identified as the best inhibitors of SARS-CoV-2 main protease. Lindane performed slightly

better than the other two ligand molecules in different post- screening studies. This study recommends that Lindane, Gluconolactone, and Mitoxantrone be the best SMP inhibitors that could be directed against SARS-CoV-2 infection. These compounds should also work against SARS coronavirus main protease since both the proteases are structurally almost identical (Figure 1).

However, computational exploration is mostly based on the modeling of the molecules and sometimes may come out with faulty outcomes, although the growing techniques have increased its fidelity of prediction, and these techniques have become famous for computeraided drug designing in the last few decades [76]. So, further laboratory experiments might be required to strengthen the findings of this study.

CONCLUSION

The Wuhan Novel Coronavirus outbreak in China caused many deaths worldwide, and many people got infected just after few days of the outbreak. Finding a cure for this nasty virus has become the primary concern of the scientific community. Our study screened a total of 1615 FDA- approved structures against SARS-CoV-2 main protease and gradually explored Lindane, Gluconolactone, and Mitoxantrone as the best inhibitors of this enzyme. These compounds were then subjected to evaluation in different drug-like parameter defining experiments where they were also performing sound. The authors believe this study will uphold the scientists' efforts to find a cure against SARS-CoV-2 infection. However, the authors suggest further in vivo and in vitro experiments for proper validation of this experiment.

CONFLICT OF INTEREST

The authors declare no conflict of interest regarding the publication of the manuscript.

DATA AVAILABILITY STATEMENT

The authors made all the data generated during the experiment and analysis available within the manuscript.

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